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Address for correspondence and reprints: Dr. Christine Gicquel, Laboratoire d'Explorations Fonctionnelles Endocriniennes, Hôpital Trousseau, 26 Avenue Arnold Netter, 75012 Paris, France. E-mail: christine.gicquel@trs.ap-hop-paris.fr

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To Trust or Not to Trust an Idiosyncratic Mitochondrial Data Set

To the Editor:

In a recent report, Silva et al. (2002) provided partial (8.8 kb) information on the mtDNA coding region (within the region 7148–15946, in the numbering of the Cambridge reference sequence [CRS]; Anderson et al. [1981]) in 40 individuals from Brazil. On the basis of the similarity in nucleotide diversity and age estimates of the four founder haplogroups A, B, C, and D, they claimed to have added new evidence for a single early entry of the founder populations into America. However, a site-by-site audit of the data reveals that their sequences are not of high enough quality to justify such statements. The authors failed to realize that a large number of mutations associated with basal branches of the worldwide mtDNA phylogeny (Finnilä et al. 2001; Maca-Meyer et al. 2001; Torroni et al. 2001; Derbeneva et al. 2002; Herrnstadt et al. 2002; Kivisild et al. 2002) were not correctly scored in their data set.

Table 1

Sequence Variation in 40 Samples Reported by Silva et al. (2002)

Sample ID	Haplogroup	Sequence on Region 7148–15976[a]	Basal Mutations Missed[b]	Accession Number
GRC0149	A	7369 7522G 8027 **8794** 8860 *11335* 12007 12705 15326 15524	11719	AF465949
KTN0130	A	**8794** 8860 11129 11288 11719 12007 12406 12705 14178 14755 14861	15326	AF465956
KPO0013	A	8764 **8794** 9392 9966 *11335* 11719 12007 12292G 12314G 12705 13708 *14566*	8860 15326	AF465957
PTJ0003	A	**8794** 11719 *11944* 12007 12705	8860 15326	AF465962
WTE1182	A	**8794** 8860 11617G 11719 12007 12292G 12618 12705 15326	...	AF465972
WPI0167	A	**8794** 8860 *10398* 11719 12007 12705 14978	15326	AF465974
YAN0623	A	**8794** 8860 10694 11719 12007 12705 13928C 15317 15326	...	AF465975
YAN0665	A	**8794** 8860 11719 12007 12705 13928C	15326	AF465976
KCR0029	A	**8794** 8860 9192 *10398* ***10400*** *11335* 12007 12314G 12705	11719 15326	AF465950
GRC0169	B4b	7626 8860 9950 *11335* 11719 11821 **13590** 15326 **15535**	8281–8289del	AF465953
KTN0209	B4b	8860 11150 11719 **13590** 14645 14647 **15535** 15914C	8281–8289del 15326	AF465955
KPO0001	B4b	7369 7522G **8281–8289del** 8860 9950 *11335* 11719 **13590** **15535**	15326	AF465958
KPO0039	B4b	8736 8860 9950 10954 *11335* 11719 **13590** **15535**	8281–8289del 15326	AF465959
KPO0023	B4b	8552 10604 11719 **13590** 13708 **15535**	8281–8289del 8860 15326	AF465960
QUE1876	B4b	8020 8860 *11335* 11719 12618 **13590**	8281–8289del 15326 15535	AF465964
QUE1881	B4b	8860 9950 *11335* 11719 **13590** *15043* **15535**	8281–8289del 15326	AF465965
YAN0637	B4b	8860 9950 11177 11719 12155C **13590** 13708 15106 **15535**	8281–8289del 15326	AF465980
KRC0033	B4b	7227T 7251 8860 9950 *10398* ***10400*** *11335* 11719 **13590**	8281–8289del 15535 15326	AF465951
QUE1880	B4b	7231C 8860 9950 *10398* ***10400*** *11335* 11719 12192 **13590** 15326	8281–8289del 15535	AF465968
JAP1044	B4c/B4a	*10115* **10238***del* 10398 *11335* 11719 15326 **15346**	8281–8289del 8860	AF465948
ARL0058	C	**7196A** 8078 **8584** 8701 9540 **9545** 10398 10400 10873 11719 **11914** 12705 **13263** 14783 15043 15301 15326	8860 14318 15487T	AF465945
PTJ0068	C	8701 8860 9540 **9545** 10873 11719 **11914** 12705 **13263** **14318** 14783 14788 15043 15914C	7196A 8584 10398 10400 15301 15487T 15326	AF465961
QUE1875	C	**7196A** **8584** 8701 8860 9540 **9545** *11335* 11719 **11914** 12705 **13263** 13656 14783 15043 15301	10398 10400 10873 14318 15487T 15326	AF465966
QUE1878	C	**8584** 8701 9540 **9545** 10873 *11335* 11719 **11914** 12705 **13263** 13545 14783 15043 15191	7196A 8860 10398 10400 14318 15301 15487T 15326	AF465967
YAN0669	C	8701 8848 8860 9540 **9545** 10310 10398 10400 11719 **11914** 12705 **13263** 13326 **14318** 14783 15043 15326	7196A 8584 10873 15301 15487T	AF465977
YAN0591	C	**8584** 8701 8848 8860 9540 **9545** 10873 11719 **11914** 12705 **13263** 13326 14783 15043	7196A 10398 10400 14318 15301 15487T 15326	AF465978

YAN0650	C	**7196A** 8701 8848 8860 9540 **9545** 10398 10873 11617G 11719 **11914** 12705 **13263** 13326 **14318** 14783 15043 15301	8584 10400 15487T 15326	AF465979
JAP1045	D4	8701 8860 8964 9296 9540 9824A *10115* 10398 10873 11719 12705 14783 15043 15301 15326	8414 10400 14668	AF465947
GRC0131	D4	8701 8860 9540 10816T 10873 *11335* 11914 12705 13059 13067 14783 15043	8414 10398 10400 11719 14668 15301 15326	AF465952
JAP1043	D4	8701 8860 9540 10398 10400 10873 11215 11719 12705 14783 15043 15301 15326 15874	8414 14668	AF465946
KTN0018	D	8701 8860 9540 10873 10874 12705 14687 14783 15043	10398 10400 11719 15301 15326	AF465954
PTJ0001	D	8701 8860 9540 10398 10400 10873 11150 11719 12705 14783 15043 15106 15301	15326	AF465963
TYR0004	D	8701 8860 9540 10398 10400 11719 12406 12705 12810 15301	10873 14783 15043 15326	AF465969
TYR0016	D	8701 8860 10398 10400 10819 10873 10874 11719 12406 12705 12810	9540 14783 15043 15301 15326	AF465970
NGR0524	L2a	**7175** 7256 **7274** 7521 *8047del* 8701 8860 **9221** 9540 **10115** 10398 10873 11719 **11914 11944** 12314G **12693** 12705 **13590** 13650	7771 8206 13803 14566 15301 15326 15784	AF465941
NGR0522	L2a	7256 **7274** 7521 **7771** 8701 8860 **9221** 9540 10873 10994C 11029T *11335* 11719 **11914 11944** 12292G **12693** 12705 **13590** 13650 **13803 15784** *15802del 15848del*	7175 8206 10115 10398 14566 15301 15326	AF465942
NGR0475	L2a	**7175** 7256 **7274** 7521 **7771** 8701 8860 **9221** 9540 10373 10873 11719 **11914 11944 12693** 12705 **13590** 13650 **13803** *14668* **15784**	8206 10115 10398 14566 15301 15326	AF465943
NGR0510	L2a	7256 **7274** 7521 **7771** 8701 8860 **9221** 9540 **10115** 10398 10873 11617G 11719 **11914 11944 12693** 12705 **13590** 13650 **13803 15784**	7175 8206 14566 15301 15326	AF465944
WTE1150	L2a	**7175** 7256 **7274** 7521 **7771** 8701 8860 **9221 10115** 10398 10873 *11335* 11719 **11914 11944 12693** 12705 13194 **13590** 13650 **13803** 15301 15326 **15784**	8206 9540 14566	AF465973
WTE1145	U	7220A 7227T 7642 8860 9668 **11467** 11719 **12308 12372** *13590* 15326	...	AF465971

NOTE.—Sites are numbered according to the revised reference sequence (Andrews et al. 1999); suffixes A, G, C, and T indicate transversions; "del" indicates a deletion. The mutations in boldface distinguish each sequence from the nearest mtDNA ancestor of haplogroups L2/3, M, N, and R. Potential reading errors or possible phantom mutations are italicized and underlined.

[a] All bear 14766 in addition.

[b] Basal polymorphisms that were undetected or omitted by Silva et al. (2002), including 11719 and the two rare mutations (8860 and 15326) in the CRS.

In the case of the hypervariable segments of the mtDNA control region, Bandelt et al. (2001, 2002) have highlighted lab-specific idiosyncrasies through comparative phylogenetic analysis. For the coding region, the task of identifying anomalies and reconstructing their potential causes is somewhat easier because the vast majority of sites there do not appear to undergo frequent mutations. The coding region well supports a basal nesting of (monophyletic) haplogroups, many of which had already been identified through RFLP analysis and sequencing of the hypervariable segments (Richards and Macaulay 2001). For example, the basal division of Eurasian mtDNAs into macrohaplogroups M and N is amazingly clear cut. The Eurasian mtDNA phylogeny that emerges from the phylogenetic analysis of the complete mtDNA database is detailed (for east Asia) in figure 1 of Kivisild et al. (2002), which attempts a reconstruction of the mutational history. The African mtDNA phylogeny has also been well documented in recent papers (Maca-Meyer et al. 2001; Torroni et al. 2001; Herrnstadt et al. 2002).

Silva et al. (2002) reported 40 mtDNAs, of which they assigned 31 to the Native American haplogroups A, B, C, and D (according to their fig. 1). The remaining nine mtDNAs can be assigned unambiguously to the Asian haplogroups B4 and D4, the Eurasian haplogroup U, and the African haplogroup L2a (table 1), as we will argue below. Figure 1 displays the truncation (relative to the 8.8-kb fragment under study) of the rooted phylogeny that is relevant for assigning these 40 mtDNAs to their respective haplogroups. This phylogeny is unanimously supported by the earlier publications. (However, note that mutations at 15301 and 11944 were not reconstructed most parsimoniously along the African mtDNA tree shown in fig. 1 of Herrnstadt et al. [2002]). The only instances of recurrent mutations (real or not) for the mutations and haplogroups highlighted in figure 1 are then as follows: the transversion 15487T is missing in the single haplogroup C lineage of Maca-Meyer et al. (2001); in the data of Herrnstadt et al. (2002), the B4b lineage 375 has experienced a transition at 14766, the L2a lineage 223 lacks the 7521 transition, and the 14566 transition is missing in the L2a lineage 165, which is closely related to another L2a lineage (bearing the 14566 mutation) from Torroni et al. (2001) in that they both share additional mutations at 3010 and 6663.

It is conspicuous that in all five haplogroup L2a mtDNAs of Silva et al. (2002), two of the basal transitions, 8206 and 14566, characteristic of L2 and L2a, respectively, are missed. Further L2a-diagnostic mutations, such as 7175, 7771, 13803, and 15784, are not always reported in the sequences (table 1). Moreover, the five L2a lineages have a total of only 11 other (private) mutations, comprising as many as five transversions, four deletions, and only two transitions. This pattern of private mutations differs from that in the three L2a lineages (nine transitions and no other mutations) of Ingman et al. (2000) and Torroni et al. (2001) in the same mtDNA region. It thus looks as though most of the real private mutations in the L2a mtDNAs were missed and that, instead, phantom mutations were scored.

The basal mutation 15487T of haplogroup M8 (which embraces haplogroups C and Z) is omitted in all seven C lineages of Silva et al.'s data (table 1). Other basal mutations for haplogroup C lineages are missing at sites 7196A, 8584, and 14318, in different combinations. It is remarkable that even deep mutations, such as 10400, 10873, and 15301 that distinguish macrohaplogroups M and N, were overlooked in six of the seven C lineages.

Among the seven D lineages in Silva et al. (2002), three sequences share mutations or motifs with D sequences reported elsewhere (Ingman et al. 2000; Derbeneva et al. 2002). The sequence JAP1045 (from an individual of Japanese origin) shares 8964, 9296, and 9824A with a Japanese mtDNA sequence from Ingman et al. (2000) and, therefore, definitely belongs to haplogroup D4, although the two characteristic D4 transitions (8414 and 14668) are not reported in the entire data set, except for one occurrence of 14668 in an L2a sequence! Similarly, the Japanese mtDNA sequence JAP1043 bears one of the mutations, 11215, found in Siberian mtDNAs of haplogroup D4 (Ingman et al. 2000; Derbeneva et al. 2002). The Guarani sequence GRC0131 of Silva et al. (2002) shares a rare transversion 10816T and a rare transition 13059 with the Guarani sequence of Ingman et al. (2000), but only the latter one has 8414 and 14668 and is thus confirmed as belonging to D4. These cases provide strong evidence for the systematic oversight of the basal mutations 8414 and 14668 in all haplogroup D lineages from Silva et al. (2002). Just as in the case of haplogroup C, several of the basal mutations that separate M and N are also missing in most of the D lineages.

Anomalies are also found in the nine sequences belonging to haplogroup A, although it was claimed by Silva et al. (2002) to be "the most homogeneous and best characterized" cluster in figure 1. Sample KCR0029 contains basal mutations 10398 and 10400 for haplogroup M. Sample KPO0013 has the 14566 mutation that is characteristic of haplogroup L2a. Sample PTJ0003 bears the L2abc-specific mutation 11944. Moreover, site 8027 is found mutated in only one A lineage, whereas this mutation was present in all the A sequences in Herrnstadt et al. (2002) and in one Chukchi sequence reported by Ingman et al. (2000).

In the 11 B lineages, only sample KPO0001 has the 9-bp deletion in the COII/tRNA^{Lys} intergenic region, characteristic of haplogroup B. One or both of the basal mutations of B4b, 13590 and 15535, occur in all the samples (with the exception of JAP1044) and hint that they belong to B4b. It should be noted that in Herrnstadt

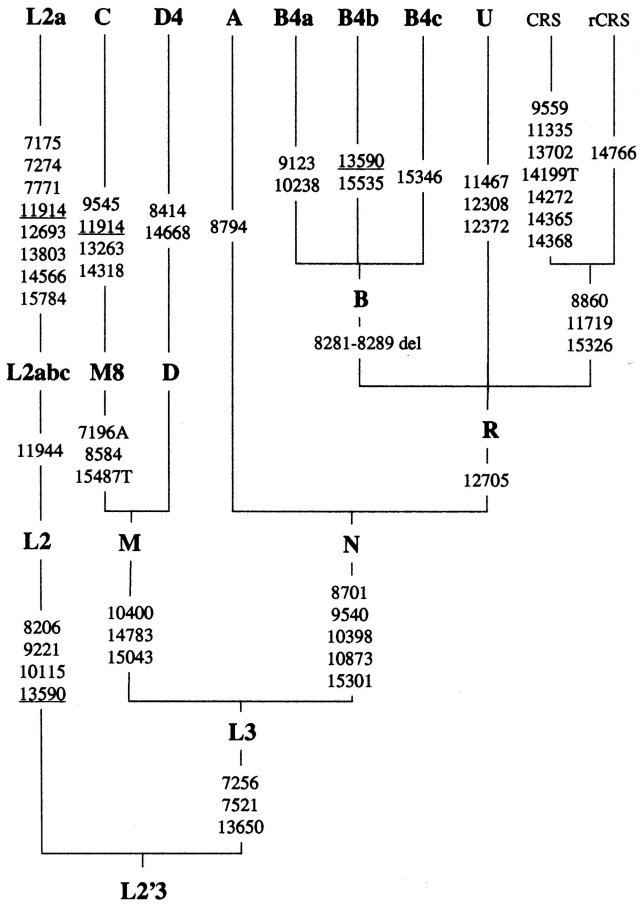

Figure 1 Skeleton of the basal mtDNA phylogeny for the haplogroups identified in the data of Silva et al. (2002). "CRS" and "rCRS" refer to the reference sequence of Anderson et al. (1981) and the revised reference sequence of Andrews et al. (1999), respectively. The suffixes A, G, C, and T indicate transversions, and "del" indicates a deletion. Parallel mutations in different branches are underlined.

et al. (2002), mutations 9950 and 11177 further defined a subhaplogroup of B4b that was baptized "B2." We suggest that the 11177 mutation could have been omitted by Silva et al. (2002) as well. The Japanese B lineage JAP1044 could belong to haplogroup B4c or, alternatively, to B4a, as judged by the 15346 mutation or the 10238 transition, respectively (if the latter was simply misreported as a deletion). Two samples, KRC0033 and QUE1880, bear the 10400 mutation of haplogroup M, whereas sample QUE1881 harbors the 15043 mutation of M.

The U sequence in Silva et al. (2002) contains the full motif of haplogroup U, plus two transversions and three transitions not previously found in the published U sequences (Ingman et al. 2000; Finnilä et al. 2001; Maca-Meyer et al. 2001; Herrnstadt et al. 2002).

Rare deletions are found in two L2a and one B lineage of Silva et al. (2002). The 15802delA and 15848delA in the cytochrome *b* gene of sample NGR0522, 8047delT in the COII gene of sample NGR0524, and 10238delT in the ND3 gene of sample JAP1044 generate premature stop codons in these genes. These rare deletions all occur at a 2-bp repeat of the deleted base and might be generated by the Sequencer reading program. It is clear that the sequences of Silva et al. (2002) harbor more rare transversions and fewer private transitions than other reported sequences (Ingman et al. 2000; Finnilä et al. 2001; Maca-Mayer et al. 2001; Torroni et al. 2001; Herrnstadt et al. 2002). One cannot exclude the possibility that true transitions were erroneously scored as transversions or deletions by Silva et al. (2002). The two rare mutations 8860 and 15326 of the CRS are also missed in most of the sequences. The mutation 11335 in the CRS, which was found to be a sequencing error (Andrews et al. 1999), was present in 16 mtDNAs.

Processes that could account for these anomalies include the following:

1. Only one strand of mtDNA was sequenced;
2. Sequences were aligned with some variant of the CRS (a likely source of problems in the past; see Macaulay et al. [1999]);
3. Sequences from different samples, especially those belonging to different haplogroups, were aligned together during the editing process (In this way, one might easily "borrow" a fragment of one sample into another when the sequences of the latter were not overlapping and, thus, introduce basal polymorphisms of one mtDNA lineage into another);
4. Possible sample crossover or contamination during data collection;
5. Relying just on the sequence scored by the Sequencer reading program without further manual checking of the chromatogram, especially relevant in the case of the rare deletions; and/or
6. PCR errors during amplification.

In summary, we have every reason to mistrust the mtDNA sequences published by Silva et al. (2002). One cannot escape the conclusion that these data are seriously flawed or, at least, are not mtDNA as we know it.

YONG-GANG YAO,[1] VINCENT MACAULAY,[3] TOOMAS KIVISILD,[4] YA-PING ZHANG,[1,2] AND HANS-JÜRGEN BANDELT[5]

[1]*Kunming Institute of Zoology, Chinese Academy of Sciences, and* [2]*Laboratory for Conservation and Utilization of Bio-Resource, Yunnan University, Kunming, Yunnan, China;* [3]*Department of Statistics, University of Oxford, Oxford, United Kingdom;* [4]*Institute of Molecular and Cell Biology, Tartu University, Tartu, Estonia; and* [5]*Fachbereich Mathematik, Universität Hamburg, Hamburg*

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Address for correspondence and reprints: Dr. Yong-Gang Yao, Kunming Institute of Zoology, Chinese Academy of Sciences, 32 Jiaochang Donglu, Kunming, Yunnan, 650223, China. E-mail: ygyaozh@yahoo.com

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Correction: Mitochondrial DNA Variation in Amerindians

To the Editor:

We thank Yao et al. (2003 [in this issue]) for calling our attention to inconsistencies in our data reporting mitochondrial DNA variations in Amerindians (Silva et al. 2002). We reviewed the original chromatograms and resequenced all the samples (forward and reverse). On the basis of the reanalysis of the initial data and sequencing that has been repeated, we conclude that most criticisms of Yao et al. are correct. We identified two sources of problems: (*a*) alignment with a variant CRS (Macaulay et al. 1999) and (*b*) mutations missed at regions of low-quality chromatograms in one (forward or reverse) of the first sequencing. Elimination of these two problems, by a second (and, in a few cases, a third) sequencing, careful manual checking of the chromatograms, and use of the correct rCRS reference sequence (MITOMAP) eliminated the discrepancies. A summary of all 40 corrected sequences is presented in figure 1, and the general pattern is similar to that recently reported by Herrnstadt et al. (2002). The presence of a private mutation in more than one individual or the absence of a basal mutation probably represent examples of homoplasy or of reverse mutations. Extensive homoplasy within the coding region of mtDNA has been documented (Eyre-Walker et al. 1999; Herrnstadt et al. 2002) and will probably be found more often as the number of mtDNA samples sequenced increases. For instance, the group C basal mutation 9545G was found in one individual from the haplogroup A, whereas private mutation 14460G was found in two individuals who belong to haplogroups A and D, and 15670C is present in one individual who belongs to haplogroup A and two who belong to haplogroup C (Herrnstadt et al. 2002). The finding of two similar private mutations (12406A) in two individuals of the same tribe (TYR0004 and TYR0016) is probably the consequence of a single mutational event, as is the occurrence of the reverse mutation 8584 in two individuals of another tribe (YAN0669 and YAN0650).

Recalculation of the age estimates for the four founder haplogroups on the basis of the reviewed data continues